



■ REVIEW ARTICLE

The pathophysiology of disc degeneration

A CRITICAL REVIEW

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The pathophysiology of intervertebral disc degeneration has been extensively studied. Various factors have been suggested as influencing its aetiology, including mechanical factors, such as compressive loading, shear stress and vibration, as well as ageing, genetic, systemic and toxic factors, which can lead to degeneration of the disc through biochemical reactions. How are these factors linked? What is their individual importance? There is no clear evidence indicating whether ageing in the presence of repetitive injury or repetitive injury in the absence of ageing plays a greater role in the degenerative process. Mechanical factors can trigger biochemical reactions which, in turn, may promote the normal biological changes of ageing, which can also be accelerated by genetic factors. Degradation of the molecular structure of the disc during ageing renders it more susceptible to superimposed mechanical injuries.

This review supports the theory that degeneration of the disc has a complex multifactorial aetiology. Which factors initiate the events in the degenerative cascade is a question that remains unanswered, but most evidence points to an age-related process influenced primarily by mechanical and genetic factors.

Disc degeneration is common, but a universally accepted definition has proved elusive. For surgeons and radiologists, degeneration might mean the presence of osteophytes and loss of signal intensity on MRI. To a biochemist, it may be expressed by changes in the content of proteoglycans or water. To a pathologist, the disc is dry, with cracks and fissures. The reason for this variance is that different disciplines use different tools, and hence see different things. This review attempts to provide a unified model of disc degeneration, bringing together all the known facts and accommodating many of the conjectures about its nature. It attempts to redress some of the misconceptions. To what degree the process of disc degeneration can contribute to back pain and disc herniation are outside the scope of this paper.

A thorough review of the literature up to January 2008 was carried out using electronic and manual searches. We retrieved and reviewed 3212 papers. Only 169 original articles addressing the pathology of disc degeneration were deemed pertinent.

The normal disc

At a molecular and cellular level, the lumbar intervertebral disc has similar constituents to

articular cartilage. Chondrocyte-like cells synthesise type II collagen, proteoglycans, and non-collagenous proteins that collectively form the matrix of the nucleus pulposus and the cartilaginous vertebral endplate. Fibroblast-like cells synthesise type I and type II collagen for the annulus fibrosus (Fig. 1). Proteoglycans consist of a core protein from which radiate chains of glycosaminoglycans containing keratan sulphate and chondroitin sulphate. Multiple proteoglycans are joined to a hyaluronic acid chain to form aggregates. Aggregates are held together by type II collagen, which is cross-linked by type IX collagen.¹ The hydroscopic properties of the proteoglycan matrix endow the nucleus with hydrostatic properties,² allowing it to accommodate compression loads and to brace the annulus. However, the constituents of the matrix are not static. They are continually degraded by enzymes, the matrix metalloproteinases (MMPs), which are secreted by the chondrocytes.³⁻⁵ Degradation of the matrix allows it to be refreshed by newly-synthesised components.

Growth factors, such as basic fibroblast growth factor (bFGF), transforming growth factor (TGF) and insulin-like growth factor (IGF), stimulate the chondrocyte or fibroblast

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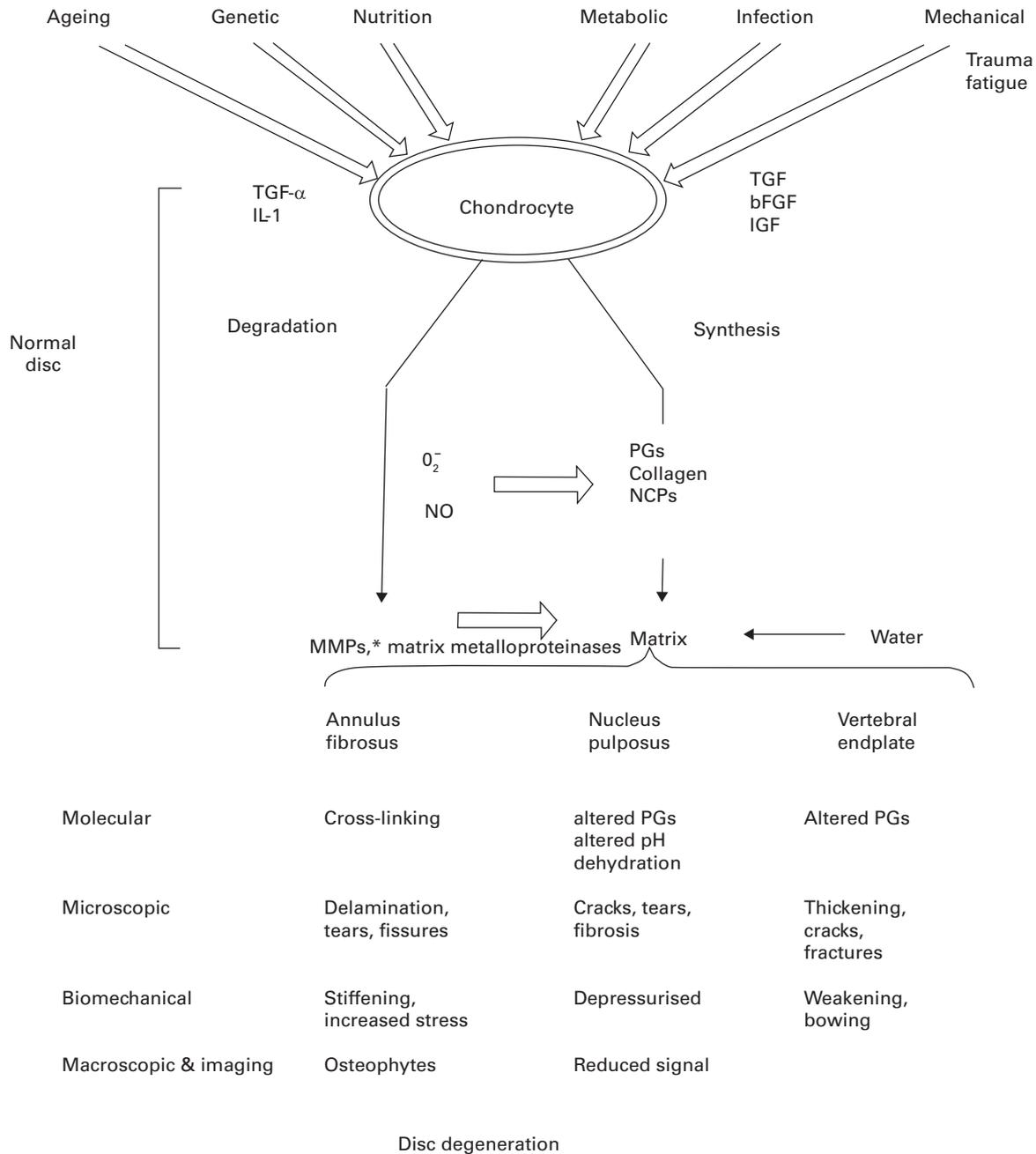


Fig. 1

A model showing disc degeneration. In a normal disc, the chondrocytes subserve a balance between the synthesis and degradation of the matrix. Synthesis is promoted by growth factors such as transforming growth factor (TGF), basic fibroblast growth factor (bFGF), and insulin-like growth factor (IGF). Degradation is activated by tumour necrosis factor- α (TNF- α) and interleukin-1 (IL-1), supplemented by superoxide (O_2^-) and nitric oxide (NO). This balance can be disturbed in favour of degradation by a number of aetiological factors. That degradation can be expressed by various features at molecular, microscopic, biomechanical, and macroscopic levels, and some can be demonstrated by medical imaging (*MMPs, matrix metalloproteinases).

to produce more matrix, and inhibit the production of MMPs.⁶ These growth factors are normally bound by cartilage intermediate layer protein (CLIP)⁶ and are released if the matrix is degraded, in order to promote further synthesis. Tissue inhibitors of metalloproteinases (TIMP) suppress the activation of MMPs, thereby controlling degradation.

Decreases in pH lessen the rate of synthesis of matrix proteoglycans.⁷

Cytokines, such as interleukin-1 (IL-1), interferon (IFN), and tumour necrosis factor- α (TNF- α) inhibit the synthesis of the matrix and promote the production of MMPs.⁸ These cytokines are produced by macrophages which enter the disc

in response to injury.⁹ Macrophages also secrete superoxide (O_2^-), which can degrade hyaluronic acid and proteoglycans, causing them to deaggregate, and can inhibit chondrocyte proliferation and synthesis. TNF- α and IL-1 stimulate inducible nitric oxide synthetase to produce nitric oxide,¹⁰ which has a variety of degradative effects. It affects matrix constituents directly, inhibits TIMPs, and thereby promotes matrix degradation and inhibits matrix synthesis.^{10,11}

Degeneration

Disc degeneration will occur if the matrix is not normal. This can arise if the components synthesised are themselves abnormal, or if the balance between synthesis and degradation of normal components is disturbed in favour of degradation. At a molecular level, degeneration will be expressed by the production of abnormal components of the matrix or by an increase in the mediators of matrix degradation (IL-1, TNF- α , superoxide and nitric oxide)¹⁰ and of MMPs and a reduction in the levels of TIMPs.^{12,13} Meanwhile, increased concentrations of growth factors (bFGF, TGF) will reflect attempts to repair the degraded matrix.^{14,15} At macroscopic and microscopic levels degeneration will become evident in the form of structural changes and defects caused by the altered matrix or impaired function of the disc (Fig. 1).

Several factors have been implicated or postulated as causing disc degeneration.

Ageing. Changes occur as the disc ages. Individually and collectively these changes reflect impaired synthesis of the matrix. The concentration of cells in the disc declines with age, especially in the annulus.¹⁶⁻¹⁸ The rate of synthesis of proteoglycans decreases,¹⁹ as does the concentration of proteoglycans in the nucleus.²⁰⁻²³ The proteoglycans produced are smaller²⁴⁻²⁶ and less aggregated¹⁹ because of a decline in link proteins²¹ and type IX collagen.^{27,28} The concentration of chondroitin sulphate falls, resulting in a rise in the ratio of keratan sulphate to chondroitin sulphate.^{22,29,30}

The collagen content of the nucleus increases and changes from type II to type I,^{21,31-34} rendering the nucleus more fibrous.³⁵ The distinction between the nucleus and annulus becomes less apparent as the two regions coalesce.³⁶ Non-collagenous proteins in the nucleus increase.³⁷⁻⁴¹ Increased collagen and increased collagen-proteoglycan binding leave fewer polar groups of the proteoglycans available to bind water.³¹ The nucleus becomes progressively more solid, dry and granular,³⁶ and cracks appear in the desiccated, fibrous nucleus.³⁵ The collagen lamellae of the annulus increase in thickness and become increasingly fibrillated.⁴²⁻⁴⁵ Cracks and cavities may develop within it.^{21,45}

Although these changes have been extensively described, their cause is not known. Among the possibilities that have been raised are declining nutrition, cell senescence, accumulation of degraded matrix products and fatigue failure of the nucleus.²¹

Cells from the intervertebral discs are subject to senescence and lose their ability to proliferate.^{46,47} Senescent cells may induce degeneration by decreased anabolism or increased catabolism.⁴⁷⁻⁴⁹ Herniated discs are associated with an increased degree of senescence.⁴⁷ The accumulated senescent cells reduce the ability of the disc to replace those lost to necrosis or apoptosis.⁴⁶

However, some features of disc degeneration are not age-related. Narrowing of the intervertebral discs has previously been considered one of the signs of ageing of the lumbar spine,^{36,50} but post-mortem studies have shown that lumbar discs do not narrow with age,⁵¹ implying that a process other than ageing is responsible. Similarly, although pathologists regard tears of the annulus as a degenerative change,^{35,36} it has been shown that radial tears do not correlate with age.⁵² Such fissures are indicative of another process. Furthermore, although all discs are the same age, those at lower lumbar levels exhibit degenerative changes far more often than in the upper levels.⁵¹ This indicates mechanical loading as the causative factor, rather than simply ageing.

Genetic factors. Studies in twins have shown a genetic predisposition to disc degeneration.⁵³⁻⁵⁸ Environmental factors only have a modest effect in identical twins.⁵⁵ These population studies have prompted searches for how genetic factors might cause degeneration. A few possible mechanisms have been identified, particularly genes encoding molecules related to the properties of the extracellular matrix. Taq I and Fok I of the vitamin D receptor gene have been implicated in disc degeneration in several studies.^{53,59,60} Taq I increases the decay by 30% of the messenger RNA that signals vitamin D efficiency,⁶¹ which impairs the sulphation of glycosaminoglycans by vitamin D.⁶² The prevalence of the t-allele of Taq I, and hence the risk for disc degeneration, differs among races. It is present in 43% of Caucasians and 31% of Africans, but in only 8% of Asians.⁶³

Polymorphism (5A and 6A alleles) commonly occurs in the promoter region of the gene that regulates MMP-3 production.⁶⁴ The 5A allele is a possible risk factor for accelerated degenerative changes of lumbar discs in the elderly, but not in the young.⁶⁵

Trp2 and Trp3 alleles result in the substitution by tryptophan of the amino acids in type IX collagen encoded by COL9A2 and COL9A3 genes. Patients with these variants of type IX collagen have an increased risk for lumbar disc disease and chronic sciatica.^{66,67} The tryptophan-containing type IX collagen gene probably produces an unstable triple helix that increases the susceptibility of disc tissue to mechanical stress.⁶⁸ Those who carry the Trp3 allele are at an increased risk of developing disc degeneration if they are obese,⁶⁹ implicating an interplay between genes and environmental factors. The prevalence of Trp alleles and their relevance as risk factors varies with ethnicity. Trp3 occurs in 24% of Finnish patients with disc degeneration,⁶⁶ but is absent in the southern Chinese.⁷⁰ Conversely, Trp2 occurs in 20% of southern Chinese

patients with disc degeneration,⁷⁰ but in only 4% of Finnish patients suffering from sciatica.⁶⁷ Meanwhile, studies in Greek populations have refuted Trp2 and Trp3 alleles as major risk factors for disc degeneration.⁷¹

Chondroitin sulphate (CS) chains are present in two adjacent regions of the proteoglycan molecule: the CS1 and CS2 domains. In the human AGCI gene, the region coding for the CS1 domain exhibits specific polymorphism⁷² which is associated with disc degeneration.⁷³ The prevalence of this gene in populations has not been determined.

A single nucleotide polymorphism (1184C allele) results in an amino acid substitution in the CLIP gene that encodes cartilage intermediate layer protein. The presence of an 1184C allele increases the binding and inhibition of growth factors such as TGF- β , reducing the growth factor-mediated induction of matrix formation genes.⁶

Among the genes that code for interleukin-1, the alleles (IL-1 α T⁸⁸⁹ and IL-1 β T³⁹⁵⁴) are associated with disc bulging, with odds ratios of 2.4 and 3.0, respectively, suggesting a genetic predisposition to disc degeneration through alterations in the function of pro-inflammatory mediators.⁷⁴

Nutrition. The intervertebral disc is the largest avascular tissue in the body. Cells in the centre of an adult lumbar disc are approximately 8 mm away from the nearest blood supply.⁷⁵ Cells in the outer annulus obtain nutrients from blood vessels in the soft tissues around its periphery, and from a sparse penetration of capillaries into its outermost region.⁷⁶ The nucleus and the cells of the inner annulus rely on a more complicated path extending from the blood vessels of the vertebral body to a capillary network that penetrates the subchondral plate.⁷⁷⁻⁷⁹ Nutrients diffuse from these capillaries across the cartilaginous endplate and through the dense disc matrix to the cells.⁸⁰ One reason for degeneration is a reduction in the transport of nutrients into the disc.⁸¹

Epidemiological and post-mortem angiographic studies indicate that insufficient blood supply to the lumbar spine due to atheromatous lesions in the abdominal aorta, or congenital hypoplasia of the lumbar arteries, could be a causative factor in disc degeneration.⁸²⁻⁸⁵ The capillary network at the bone-cartilage endplate junction diminishes after the first decade of life when the first signs of disc degeneration become evident.^{18,76,86} Calcification of the endplate occludes the vascular openings within it, acting as a barrier to transport of nutrients because of lowering of endplate permeability.^{18,76,80,81,87-89} Calcification of the endplate in scoliotic patients has been correlated with loss of nutrient and cell death.^{90,91} The capillary network is regulated by vasoactive agents such as noradrenaline (norepinephrine) and acetylcholine,⁹² and by mechanical stimuli such as vibration.^{93,94} Smoking is associated with an increased incidence of disc herniation,⁹⁵ and degeneration⁹⁶ implying a systemic effect by constriction of arterioles or anoxia to cells in the disc induced by carboxyhaemoglobin.⁹⁷ In animal models, remodelling of

this capillary bed has been demonstrated in response to nicotine.⁹⁸

The disc relies on diffusion for its nutrition, and pivotal to this is movement, which pumps water and nutrients into the disc. Accordingly, some investigators have considered that sustained compression,^{99,100} or immobilisation even without compression,¹⁰⁰ might be the basis of impaired nutrition to the disc. However, the laboratory data are conflicting. Studies in rabbits¹⁰¹ and mice^{99,100} have found that constant loading in compression resulted in disc degeneration, but a study in dogs found that high, static compressive forces across lumbar discs for up to a year did not produce annular fissures, bulging or any other visible form of disc degeneration.^{102,103} It has also been shown that there is reduced synthesis of proteoglycans and an increase in MMP-3 after the application of pressures above and below the physiological levels.¹⁰⁴ These data are of limited clinical relevance because human subjects are unlikely to suffer prolonged static loads.

A reduced supply of nutrients leads to an increase in oxidative stress markers, seen initially in the nucleus of young discs, and eventually throughout the inner regions of the disc with increased age and degeneration.¹⁸ Deposition of carboxymethyllysine reflects a distinct cellular reaction to increased oxidative stress, and possibly impaired function.¹⁸ A low level of oxygen and an acidic pH from the anaerobic metabolism lead to a fall in protein and synthesis of proteoglycan.^{105,106} A fall in nutrient supply can also reduce the number of viable cells in the disc.¹⁰⁷

Toxic factors. Nicotine directly inhibits proliferation of disc cells and their synthesis of extracellular matrix, as shown on bovine cells from the nucleus pulposus cultured *in vitro*.¹⁰⁸ Similarly, passive smoking in rats resulted in downregulation of collagen genes which preceded the histological changes of degeneration.¹⁰⁹

Metabolic disorders. Various metabolic disorders can cause disc degeneration either by interfering with the normal biochemistry of matrix synthesis or by deposition of foreign materials in the disc.

In patients with diabetes mellitus, the nucleus pulposus demonstrates a significant decrease in hexosamine content, an increase in hydroxyproline and enhanced activity of enzymes involved in the metabolism of carbohydrates.¹¹⁰ The discs exhibit deficiencies in incorporation of ³⁵S-sulphate during proteoglycan synthesis, which indicates a reduced rate of glycosylation and a decrease in the number of sugar side chains per core protein.¹¹¹

Patients with alkaptonuria develop intradiscal deposits of a black pigment, inducing disc degeneration characterised by calcification appearing as elliptical opaque wafers.^{94,95}

Low-grade infection. It has been proposed that disc degeneration could be caused by low-grade infection. One group found that 31% of discs harvested after microdiscectomy tested positive for low-virulence Gram-positive bacteria, and 84% of these were infected with *Propionibacterium*

acnes.¹¹² Fritzell et al,¹¹³ could not confirm these findings, although two of the ten patients they studied exhibited evidence of bacterial infection unrelated to *P. acnes*.

Patients who recover poorly after surgery for disc herniation have exhibited elevated serum concentrations of high-sensitivity C-reactive protein (hs-CRP).¹¹⁴ This could be caused by interleukin-6, which occurs in degenerative discs.¹¹⁵ However, it is not clear whether CRP is a sign of disc infection or an inflammatory response to disc material in the epidural space.

Neurogenic inflammation. Upon mechanical stimulation, cells in the dorsal root ganglion produce substance P, which acts centrally as a neurotransmitter.¹¹⁶ However, some investigators have proposed that antidromic release of substance P into the disc innervated by the ganglion might produce degeneration by stimulating the synthesis of inflammatory agents and degradative enzymes in the disc.¹¹⁷ This phenomenon is unlikely to be a primary cause of degeneration because it requires a cause of irritation of the ganglion before the disc degenerates or herniates, but disc herniation is the most common cause of this. Nevertheless, it is possible that neurogenic inflammation might aggravate disc degeneration once a herniation or osteophyte irritates the ganglion.

Autoimmune theory. Damage to the endplate after a compression injury exposes the antigenic components of the nucleus pulposus to the circulation, which may trigger an immune response.¹¹⁸ This may be responsible for a chronic inflammatory reaction leading to resorption of the affected disc,¹¹⁸ or may even involve discs at different vertebral levels, resulting in multilevel degeneration.¹¹⁹ Local expression of Fas ligand by cells in the disc plays a key role in the molecular mechanism that maintains the immune-privileged characteristics of the disc by inducing apoptosis of invading Fas-positive T cells.^{120,121} Animal experiments have confirmed that violation of the physiological barrier of the disc by mechanical injury changes the role of Fas ligand and induces apoptosis of the disc cells.¹²¹

Mechanical factors. Circumstantial evidence suggests the possibility of mechanical factors in the aetiology of disc degeneration. Degeneration is more common and more severe at lower lumbar levels.⁵¹ Discs immediately above a transitional vertebra tend to exhibit significantly more degenerative changes than those between the transitional vertebra and the sacrum.¹²² Mechanical factors are postulated in the production of disc degeneration adjacent to a lumbar fusion.^{123,124}

Vibration. Vibration has been incriminated in the pathogenesis of disc degeneration. Results of animal and *in vitro* studies suggest that vibration can adversely affect the nutrition⁹⁴ and metabolism⁹³ of the disc, especially if the vibration matches the resonant frequency of the lumbar spine (4 Hz to 6 Hz).¹²⁵⁻¹²⁹ People exposed to whole-body vibration in the resonant range, such as helicopter pilots¹³⁰ and drivers of trucks, buses and tractors,¹³¹⁻¹³⁴ have a high rate of back pain. Similarly, epidemiological studies have

provided some support for the hypothesis that driving adversely affects the intervertebral discs, with a higher rate of herniation in occupational drivers.¹³⁵⁻¹³⁷ However, MRI observation in identical twins with different patterns of occupational driving do not support the hypothesis that driving is a risk factor for disc degeneration.¹³⁸ Epidemiological studies on tractor-driving farmers,¹³⁹ rally drivers,¹⁴⁰ and operators of heavy earth-moving machinery¹⁴¹ have not revealed an association between long-term exposure to vibration and disc degeneration. Back pain related to vibration exposure may be related to other factors.

Torsion. Torsional movements generate tension in half of the collagen fibres in the annulus, whereas the other fibres tend to become slack.¹⁴² For the fibres to incur damage, they must be elongated by more than 4% of their resting length,¹⁴³ and this is feasible only when axial rotation at each individual segment exceeds 3°.¹⁴⁴ For this to occur, the zygoapophyseal joints must be damaged first, as they resist rotation beyond 1.5° to 3°.^{144,145} However, when the spine is flexed, the gaping facet joints offer less constraint to rotation,¹⁴⁶ leading to an annular tear without damage to the facets.

In cadaver studies torsional injury resulted in tears at the periphery of the annulus, usually in the posterolateral region, which slowly extended radially towards the centre of the disc with further torsion.^{143,144,147,148} In the degenerative spine, the tear may start in the inner fibres of the annulus and extend toward the periphery.^{143,144,147,148} *In vivo* animal studies have confirmed that rotation of the lumbar spine after facetectomy leads to degenerative changes, with narrowing of the disc space, fissuring of the annulus and migration of the nucleus.^{149,150}

That torsion injury may lead to disc degeneration can be supported by clinical imaging studies. Professional bowlers in cricket with a chronic stress reaction of the pars interarticularis or a unilateral stress fracture tend to have an intervertebral disc of normal height and appearance on MRI. However, bilateral stress fractures are associated with severe disc degeneration.¹⁵¹ As cadaver studies have shown that the apophyseal joints resist most of the intervertebral shear,¹⁴⁶ it can be assumed that increased shear stress, presumably torsional injuries caused by unilateral pars fracture, can precipitate disc degeneration.

Compression. The lumbar disc is designed to sustain compression loads which are beneficial to the disc. Loading is the physiological stimulus for matrix turnover.^{104,152-154} It induces MMPs^{65,104,154} and nitric oxide,¹⁵⁵ ostensibly to clear the matrix in preparation for new synthesis, and induces matrix synthesis.¹⁵⁴ However, excessive loading can lead to deleterious changes in the disc by reducing gene expression of all anabolic proteins with significant effects on aggrecan formation, while simultaneously increasing gene expression of MMPs.¹⁵⁴

The most vulnerable component of a lumbar disc is the vertebral endplate. When subjected to compression, the endplate fails by fracturing. This can occur as a result of

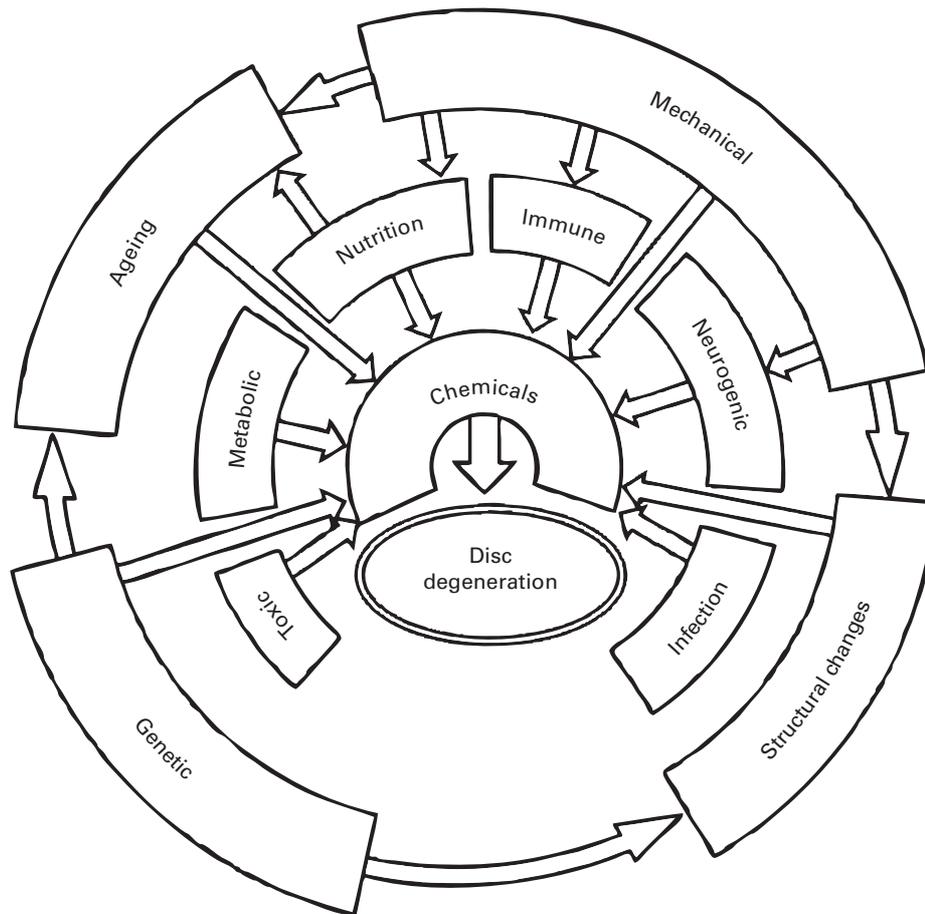


Fig. 2

A model showing the multifactorial pathophysiology of disc degeneration. The different factors may behave independently as initiators or promoters or both.

application of a sudden, severe compression load¹⁵⁶⁻¹⁵⁸ or as a result of fatigue failure.^{158,159} Repeated application of loads amounting to between 50% and 80% of the ultimate tensile strength of the endplate can cause a fracture after as few as 100 cycles.¹⁶⁰ Subsequently, endplate fractures can precipitate degeneration by a variety of mechanisms.

Callus formation might occlude blood vessels in the endplate, thereby interfering with cell nutrition and maintenance of the extracellular matrix. An *in vitro* study showed that damage to the endplate can produce adverse effects on diffusion into the disc; morphological features of degeneration correlated inversely with the density of vascular openings in the endplate.⁸⁷ Studies both in humans¹⁶¹ and in pigs¹⁶² have related the severity of disc degeneration to that of the injury to the endplate. There is a strong association between degeneration and defects in the endplate from Schmorl's nodes,¹⁶³⁻¹⁶⁵ Scheuermann's disease¹⁶⁶ and fractures,¹⁶⁷ with an increased incidence of disc prolapse, particularly at the lower lumbar levels.¹⁶⁴

Damage to the endplate rapidly leads to depressurisation of the nucleus and a simultaneous increase in stress in the

posterior annulus.^{158,159} The depressurised nucleus is no longer able to brace the annulus, as a result of which the inner lamellae of the annulus buckle inwards and the outer lamellae buckle outwards, particularly posteriorly. Finite element models have confirmed that interlaminar shear stresses arising from a compressive load are highest in the posterolateral annulus.¹⁶⁸ These stresses lead to separation of the adjacent laminae (delamination), which appears as concentric tears in the annulus fibrosus.¹⁶³

Mechanisms have been postulated whereby fractures of the endplate can lead to biochemical changes in the matrix of the disc. Mechanobiological studies both *in vivo* and *in vitro* have clearly demonstrated that compression can influence the biosynthetic activity of cells in the disc, altering the expression of key extracellular matrix molecules.^{99,169,170} Furthermore, destruction of cartilage from an endplate fracture would provoke an IL-1-mediated inflammatory response, inducing enzymes that destroy proteoglycans.⁹ If this response is extended into the adjacent matrix, it could initiate degradation.^{118,144} Another possibility is that exposure of the matrix to the blood in the vertebral body might

elicit an autoimmune response similar to that seen in sympathetic ophthalmia.¹¹⁸ A less elaborate explanation is that fractures of the endplate alter the pH of the adjacent matrix, which activates MMPs. Consistent with this view is that activation of MMPs occurs progressively away from an endplate into the nucleus.¹⁷¹

None of these mechanisms has been explicitly demonstrated, but laboratory studies have now shown that experimental injury to an endplate does produce degenerative changes.^{162,172} The water content of the nucleus decreases, the proteoglycan content decreases, the nuclear pressure falls and the inner annulus delaminates.

Discussion

Degeneration is not a diagnosis but an expression of the state of the disc, which is the result of several factors acting individually or collectively (Fig. 2). Rather than being the result of a single process, disc degeneration can have a number of possible causes. Proper diagnosis requires that the exact aetiology is clearly established.

Degeneration secondary to metabolic disorders does not pose a diagnostic problem as the general features of the condition provide the diagnosis. Where the difficulty arises is in determining whether the condition is genetic, due simply to ageing, or traumatic.

Disc degeneration can be due to genetic factors that produce abnormal components of the matrix which compromise the structure and function of the disc. However, degeneration cannot be attributed wholly to genetic factors. Epidemiological studies have shown that genetic factors increase the risk of degeneration, but they do not account for all cases, nor are they uniform across different ethnic populations. Studies across large multiethnic populations are required.¹⁷³

The classic interpretation has been that disc degeneration is a result of age-related changes. However, this interpretation is based on little more than the circumstantial evidence that the biochemical and morphological features of degeneration increase with age. What remains unexplained is why they occur. Theories of impaired nutrition have not been substantiated. The default explanation is that degeneration is due to programmed cell senescence.

An alternative view is that disc degeneration is caused by mechanical factors. Increasing evidence implicates injury to the vertebral endplate as central to the process. Such injuries can impair the nutrition of the disc or can directly precipitate matrix degeneration. The biomechanical changes of disc degeneration are as much a sign of a response by a connective tissue to injury as due to idiopathic age changes. Evidence in favour of mechanical factors is strongly based on laboratory data from cadaver experiments. It should be borne in mind that freezing animal or human cadaver specimens permanently alters disc behaviour compared with normal circumstances.^{174,175} In spite of this, reasonable evidence can be obtained to support the mechanical concept of disc degeneration, particu-

larly that originating in torsional and compressive injuries. Compressive loading or torsion may produce fracture of the endplate or tear of the annulus respectively, which, in turn, will drive the biological events.

Perhaps the most pivotal development in biomechanical research into disorders of the lumbar spine is the recognition of fatigue failure. Endplate fractures and disc degeneration do not require a single memorable traumatic event: they can occur silently and progressively as a result of repeated, subliminal insults to the disc. This renders mechanical factors difficult to identify in epidemiological studies. The failure to incriminate environmental factors in the aetiology of disc degeneration may lie in the lack of tools with which to detect them. The development of high-resolution imaging techniques may enable detection of tiny fractures of the endplate *in vivo*.

Based on the available evidence, conventional wisdom dictates that degeneration of the intervertebral disc can be defined as an age-dependent, cell-mediated molecular degradation process under genetic influence that is accelerated primarily by nutritional and mechanical factors, and secondly by toxic or metabolic influences. These factors mediate degeneration by triggering chemical reactions. These changes can affect the morphology of the disc, manifested as evidenced by thickening of the vertebral endplate, cracks and fissures in the matrix, delamination and tears in the annulus, and in the biomechanical function of the disc (Fig. 1). The end result of disc degeneration is characterised by collapse of the intervertebral space and osteophyte formation.

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References

1. **Eyre DR, Matsui Y, Wu JJ.** Collagen polymorphisms of the intervertebral disc. *Biochem Soc Trans* 2002;30:844-8.
2. **Urban JP, Maroudas A.** Swelling of the intervertebral disc in vitro. *Connect Tissue Res* 1981;9:1-10.
3. **Matrisian LM.** Metalloproteinases and their inhibitors in matrix remodeling. *Trends Genet* 1990;6:121-5.
4. **Ito A, Mukaiyama A, Itoh Y, et al.** Degradation of interleukin 1 beta by matrix metalloproteinases. *J Biol Chem* 1996;271:14657-60.
5. **Goupille P, Jayson MI, Valat JP, Freemont AJ.** Matrix metalloproteinases: the clue to intervertebral disc degeneration? *Spine* 1998;23:1612-26.
6. **Seki S, Kawaguchi Y, Chiba K, et al.** A functional SNP in CILP, encoding cartilage intermediate layer protein, is associated with susceptibility to lumbar disc disease. *Nat Genet* 2005;37:607-12.
7. **Ohshima H, Urban JPG.** The effect of lactate and pH on proteoglycan and protein synthesis rates in the intervertebral disc. *Spine* 1992;17:1079-82.
8. **Kobayashi M, Squires GR, Mousa A, et al.** Role of interleukin-1 and tumor necrosis factor alpha in matrix degradation of human osteoarthritis cartilage. *Arthritis Rheum* 2005;52:128-35.
9. **Shinmei M, Masuda K, Kikuchi T, Shimomura Y.** Interleukin-1, tumour necrosis factor, and interleukin-6 as mediators of cartilage destruction. *Semin Arthritis Rheum* 1989;18:27-32.
10. **Kang JD, Stefanovic-Racic M, McIntyre LA, Georgescu HI, Evans CH.** Toward a biochemical understanding of human intervertebral disc degeneration and herniation: contributions of nitric oxide, interleukins, prostaglandin E2, and matrix metalloproteinases. *Spine* 1997;22:1065-73.
11. **Burke JG, Watson RW, McCormack D, et al.** Intervertebral discs which cause low back pain secrete high levels of proinflammatory mediators. *J Bone Joint Surg [Br]* 2002;84-B:196-201.
12. **Kang JD, Georgescu HI, McIntyre-Larkin L, et al.** Herniated lumbar intervertebral discs spontaneously produce matrix metalloproteinases, nitric oxide, interleukin-6, and prostaglandin E2. *Spine* 1996;21:271-7.

13. Kanemoto M, Hukuda S, Komiya Y, Katsuura A, Nishioka J. Immunohistochemical study of matrix metalloproteinases-3 and tissue inhibitor of metalloproteinase-1 human intervertebral discs. *Spine* 1996;21:1-8.
14. Peng B, Hao J, Hou S, et al. Possible pathogenesis of painful intervertebral disc degeneration. *Spine* 2006;31:560-6.
15. Doita M, Kanatani H, Harada T, Mizuno K. Immunohistologic study of the ruptured intervertebral disc of the lumbar spine. *Spine* 1996;21:235-41.
16. Trout JJ, Buckwalter JA, Moore KC, Landas SK. Ultrastructure of the human intervertebral disc. I: changes in notochordal cells with age. *Tissue Cell* 1982;14:359-69.
17. Trout JJ, Buckwalter JA, Moore KC. Ultrastructure of the human intervertebral disc. II: cells of the nucleus pulposus. *Anat Rec* 1982;204:307-14.
18. Nerlich AG, Schleicher ED, Boos N. Immunohistologic markers for age-related changes of human lumbar intervertebral discs. *Spine* 1997;22:2781-95.
19. Johnstone B, Bayliss MT. The large proteoglycans of the human intervertebral disc: changes in their biosynthesis and structure with age, topography, and pathology. *Spine* 1995;20:2781-84.
20. Beard HK, Stevens RL. Biochemical changes in the intervertebral disc. In: Jayson MIV, ed. *The lumbar spine and back pain*. Second ed. London: Pitman Medical, 1980:407-36.
21. Buckwalter JA. Aging and degeneration of the human intervertebral disc. *Spine* 1995;20:1307-14.
22. Gower WE, Pedrini V. Age related variation in protein polysaccharides from human nucleus pulposus, annulus fibrosus and costal cartilage. *J Bone Joint Surg [Am]* 1969;51-A:1154-62.
23. Sylven B, Paulson S, Hirsch C, Snellman O. Biophysical and physiological investigations on cartilage and other mesenchymal tissues. II: the ultrastructure of bovine and human nuclei pulposi. *J Bone Joint Surg [Am]* 1951;33-A:333-40.
24. Adams P, Muir H. Qualitative changes with age of human lumbar disks. *Ann Rheum Dis* 1976;35:289-96.
25. Bushell GR, Ghosh P, Taylor TKF, Akeson WH. Proteoglycan chemistry of the intervertebral disks. *Clin Orthop* 1977;129:115-23.
26. Comper WD, Preston BN. Model connective tissue systems: a study of the polyonmobile ion and of excluded volume interactions of proteoglycans. *Biochem J* 1974;143:1-9.
27. Buckwalter JA, Pedrini-Mille A, Pedrini V, Tudisco C. Proteoglycans of human infant intervertebral discs: electron microscopic and biochemical studies. *J Bone Joint Surg [Am]* 1985;67-A:284-94.
28. Buckwalter JA, Woo SL, Goldberg VM, et al. Soft-tissue aging and musculoskeletal function. *J Bone Joint Surg [Am]* 1993;75-A:1533-48.
29. Adams P, Eyre DR, Muir H. Biochemical aspects of development and ageing of human lumbar intervertebral discs. *Rheumatol Rehab* 1977;16:22-9.
30. Naylor A. Intervertebral disc prolapse and degeneration: the biochemical and biophysical approach. *Spine* 1976;1:108-14.
31. Hirsch C, Paulson S, Sylven B, Snellman O. Biophysical and physiological investigation on cartilage and other mesenchymal tissues. VI: characteristics of human nuclei pulposi during aging. *Acta Orthop Scand* 1953;22:175-83.
32. Naylor A, Happey F, MacRae TP. The collagenous changes in the intervertebral disc with age and their effect on elasticity: an x-ray crystallographic study. *Brit Med J* 1954;2:570-3.
33. Naylor A, Happey F, Turner RL, et al. Enzymic and immunological activity in the intervertebral disc. *Orthop Clin North Am* 1975;6:51-8.
34. Deshmukh K, Kline WH. Characterization of collagen and its precursors synthesized by rabbit-articular-cartilage cells in various culture systems. *Eur J Biochem* 1976;69:117-23.
35. Haefeli M, Kalberer F, Saegesser D, et al. The course of macroscopic degeneration in the human lumbar intervertebral disc. *Spine* 2006;31:1522-31.
36. Vernon-Roberts B, Pirie CJ. Degenerative changes in the intervertebral discs of the lumbar spine and their sequelae. *Rheumatol Rehab* 1977;16:13-21.
37. Blakey PR, Happey F, Naylor A, Turner RL. Protein in the nucleus pulposus of the intervertebral disc. *Nature* 1962;215:52-3.
38. Dickson IR, Happey F, Pearson CH, Naylor A, Turner RL. Variations in the protein components of human intervertebral disc with age. *Nature* 1967;215:52-3.
39. Ghosh P, Bushell GR, Taylor TKF, Akeson WH. Collagens, elastin, and noncollagenous protein of the intervertebral disk. *Clin Orthop* 1977;129:124-32.
40. Naylor A. The biochemical changes in the human intervertebral disc in degeneration and nuclear prolapse. *Orthop Clin North Am* 1971;2:343-58.
41. Taylor TKF, Little K. Intercellular matrix of the intervertebral disk in ageing and in prolapse. *Nature* 1965;208:384-6.
42. Harris RI, MacNab I. Structural changes in the lumbar intervertebral discs: their relationship to low back pain and sciatica. *J Bone Joint Surg [Br]* 1954;36-B:304-22.
43. Marchand F, Ahmed AM. Investigation of the laminar structure of lumbar disc annulus fibrosus. *Spine* 1990;15:402-10.
44. Pritzker KPH. Aging and degeneration in the lumbar intervertebral disc. *Orthop Clin North Am* 1977;8:65-77.
45. Hirsch C, Schajowicz F. Studies on structural changes in the lumbar annulus fibrosus. *Acta Orthop Scand* 1953;22:184-9.
46. Gruber HE, Ingram JA, Norton HJ, Hanley EN Jr. Senescence in cells of the aging and degenerating intervertebral disc: immunolocalization of senescence-associated beta-galactosidase in human and sand rat discs. *Spine* 2007;32:321-7.
47. Roberts S, Evans EH, Kleitsas D, Jaffray DC, Eisenstein SM. Senescence in human intervertebral discs. *Eur Spine J* 2006;15(Suppl 3):312-16.
48. Oshima J, Campisi J. Fundamentals of cell proliferation: control of the cell cycle. *J Dairy Sci* 1991;74:2778-87.
49. West MD, Pereira-Smith OM, Smith JR. Replicative senescence of human skin fibroblasts correlates with a loss of regulation and overexpression of collagenase activity. *Exp Cell Res* 1989;184:138-47.
50. Lawrence JS. Disc degeneration, its frequency and relationship to symptoms. *Ann Rheum Dis* 1969;28:121-38.
51. Miller JA, Schmatz C, Schultz AB. Lumbar disc degeneration: correlation with age, sex, and spine level in 600 autopsy specimens. *Spine* 1988;13:173-8.
52. Moneta GB, Videman T, Kaivanto K, et al. Reported pain during lumbar discography as a function of anular ruptures and disc degeneration: a re-analysis of 833 discograms. *Spine* 1994;17:1968-74.
53. Videman T, Leppävuori J, Kaprio J, et al. Intragenic polymorphisms of the vitamin D receptor gene associated with intervertebral disc degeneration. *Spine* 1998;23:2477-85.
54. Battie MC, Haynor DR, Fisher LD, et al. Similarities in degenerative findings on magnetic resonance images of the lumbar spines of identical twins. *J Bone Joint Surg [Am]* 1995;77-A:1662-70.
55. Battie MC, Videman T, Gibbons LE, et al. Determinants of lumbar disc degeneration: a study relating lifetime exposures and magnetic resonance imaging findings in identical twins. *Spine* 1995;20:2601-12.
56. Matsui H, Kanamori M, Ishihara H, et al. Familial predisposition for lumbar degenerative disc disease: a case-control study. *Spine* 1998;23:1029-34.
57. Postacchini F, Lami R, Pugliese O. Familial predisposition to discogenic low-back pain: an epidemiologic and immunogenetic study. *Spine* 1988;13:1403-6.
58. Sambrook PM, MacGregor AJ, Spector TD. Genetic influences on cervical and lumbar disc degeneration: a magnetic resonance imaging study in twins. *Arthritis Rheum* 1999;42:366-72.
59. Kawaguchi Y, Kanamori M, Ishihara H, et al. The association of lumbar disc disease with vitamin-D receptor gene polymorphism. *J Bone Joint Surg [Am]* 2002;84-A:2022-8.
60. Cheung KMC, Chan D, Karppinen J, et al. Association of the Taq I allele in vitamin D receptor with degenerative disc disease and disc bulge in Chinese. *Spine* 2006;31:1143-8.
61. Fang Y, van Meurs JB, d'Alesio A, et al. Promoter and 3'-untranslated-region haplotypes in the vitamin d receptor gene predispose to osteoporotic fracture: the rotterdam study. *Am J Hum Genet* 2005;77:807-23.
62. Fernandes I, Hampson G, Cahours X, et al. Abnormal sulfate metabolism in vitamin D-deficient rats. *J Clin Invest* 1997;100:2196-203.
63. Uitterlinden AG, Fang Y, Van Meurs JB, Pols HA, Van Leeuwen JP. Genetics and biology of vitamin D receptor polymorphisms. *Gene* 2004;338:143-56.
64. Ye S, Watts GF, Mandalia S, Humphries SE, Henney AM. Preliminary report: genetic variation in the human stromelysin promoter is associated with progression of coronary atherosclerosis. *Br Heart J* 1995;73:209-15.
65. Takahashi M, Haro H, Wakabayashi Y, et al. The association of degeneration of the intervertebral disc with 5a/6a polymorphism in the promoter of the human matrix metalloproteinase-3 gene. *J Bone Joint Surg [Br]* 2001;83-B:491-5.
66. Paasilta P, Lohiniva J, Goring HH, et al. Identification of a novel common genetic risk factor for lumbar disk disease. *JAMA* 2001;285:1843-9.
67. Annunen S, Paasilta P, Lohiniva J, et al. An allele of COL9A2 associated with intervertebral disc disease. *Science* 1999;285:409-12.
68. Persikov AV, Ramshaw JA, Brodsky B. Collagen model peptides: sequence dependence of triple-helix stability. *Biopolymers* 2000;55:436-50.
69. Solovieva S, Lohiniva J, Leino-Arjas P, et al. COL9A3 gene polymorphism and obesity in intervertebral disc degeneration of the lumbar spine: evidence of gene-environment interaction. *Spine* 2002;27:2691-6.
70. Jim JJ, Naponen-Hietala N, Cheung KM, et al. The TRP2 allele of COL9A2 is an age-dependent risk factor for the development and severity of intervertebral disc degeneration. *Spine* 2004;29:1266-70.

71. Kales SN, Linos A, Chatzis C, et al. The role of collagen IX tryptophan polymorphisms in symptomatic intervertebral disc disease in Southern European patients. *Spine* 2004;29:1266-70.
72. Doege KJ, Coulter SN, Meek LM, Maslen K, Wood JG. A human-specific polymorphism in the coding region of the aggrecan gene: variable number of tandem repeats produce a range of core protein sizes in the general population. *J Biol Chem* 1997;272:13974-9.
73. Kawaguchi Y, Osada R, Kanamori M, et al. Association between an aggrecan gene polymorphism and lumbar disc degeneration. *Spine* 1999;24:2456-60.
74. Solovieva S, Kouhia S, Leino-Arjas P, et al. Interleukin 1 polymorphisms and intervertebral disc degeneration. *Epidemiology* 2004;15:626-33.
75. Katz MM, Hargens AR, Garfin SR. Intervertebral disc nutrition: diffusion versus convection. *Clin Orthop* 1986;210:243-5.
76. Boos N, Weissbach S, Rohrbach H, et al. Classification of age-related changes in lumbar intervertebral discs. *Spine* 2002;27:2631-44.
77. Holm S, Maroudas A, Urban JP, Solstam G, Nachemson A. Nutrition of the intervertebral disc: solute transport and metabolism. *Connect Tissue Res* 1981;8:101-19.
78. Crock HV, Goldwasser M. Anatomic studies of the circulation in the region of the vertebral endplate in adult greyhound dogs. *Spine* 1984;9:702-6.
79. Hassler O. The human intervertebral disc: a micro-angiographical study on its vascular supply at various ages. *Acta Orthop Scand* 1969;40:765-72.
80. Roberts S, Urban JPG, Evans H, Eisenstein SM. Transport properties of the human cartilage endplate in relation to its composition and calcification. *Spine* 1996;21:415-20.
81. Nachemson A, Lewin T, Maroudas A, Freeman MA. In vitro diffusion of dye through the end-plates and annulus fibrosus of human lumbar inter-vertebral discs. *Acta Orthop Scand* 1970;41:589-607.
82. Kauppila LI. Can low-back pain be due to lumbar-artery disease? *Lancet* 1995;346:888-9.
83. Kauppila LI, Tallroth K. Postmortem angiographic findings for arteries supplying the lumbar spine: their relationship to low-back symptoms. *J Spinal Disord* 1993;6:124-9.
84. Kauppila LI, McAlindon T, Evans S, et al. Disc degeneration/back pain and calcification of the abdominal aorta: a 25-year follow-up study in Framingham. *Spine* 1997;22:1642-9.
85. Kauppila L, Penttila A, Karhunen PJ, Lalu K, Hannikainen P. Lumbar disc degeneration and atherosclerosis of the abdominal aorta. *Spine* 1994;19:923-9.
86. Chandraraj S, Briggs CA, Opekin K. Disc herniations in the young and end-plate vascularity. *Clin Anat* 1998;11:171-6.
87. Benneker LM, Heini PF, Alini M, Anderson SE, Ito K. Vertebral endplate marrow contact channel occlusions and intervertebral disc degeneration. *Spine* 2005;30:167-73.
88. Bernick S, Cailliet R. Vertebral end-plate changes with aging of human vertebrae. *Spine* 1982;7:97-102.
89. Gruber HE, Ashraf N, Kilburn J, et al. Vertebral endplate architecture and vascularization: application of micro-computerized tomography, a vascular tracers, and immunocytochemistry in analyses of disc degeneration in the aging sand rat. *Spine* 2005;30:2593-600.
90. Urban MR, Fairbank JCT, Etherington PJ, et al. Electrochemical measurement of transport into scoliotic intervertebral discs in vivo using nitrous oxide as a tracer. *Spine* 2001;26:984-90.
91. Bibby SR, Fairbank JC, Urban MR, Urban JP. Cell viability in scoliotic discs in relation to disc deformity and nutrient levels. *Spine* 2002;27:2220-8.
92. Wallace AL, Wyatt BC, McCarth ID, Hughes SP. Humoral regulation of blood flow in the vertebral endplate. *Spine* 1994;19:1324-8.
93. Hirano N, Tsuji H, Oshihama H, et al. Analysis of rabbit intervertebral disc physiology based on water metabolism. II: changes in normal intervertebral discs under axial vibratory load. *Spine* 1988;13:1297-302.
94. Holm S, Nachemson A. Nutrition of the intervertebral disc: effects induced by vibration. *Orthop Trans* 1985;9:525.
95. Kelsey JL, Githens PB, O'Connor T, et al. Acute prolapsed lumbar intervertebral disc: an epidemiological study with special reference to driving automobiles and cigarette smoking. *Spine* 1984;9:608-13.
96. Battie MC, Videman T, Gill K, et al. Smoking and lumbar intervertebral disc degeneration: an MRI study of identical twins. *Spine* 1991;16:1015-21.
97. Holm S, Nachemson A. Nutrition of the intervertebral disc: acute effects of cigarette smoking: an experimental animal study. *Ups J Med Sci* 1988;93:91-9.
98. Iwahashi M, Matsuzaki H, Tokuhashi Y, Wakabayashi K, Uematsu Y. Mechanism of intervertebral disc degeneration caused by nicotine in rabbits to explicate intervertebral disc disorders caused by smoking. *Spine* 2002;27:1396-401.
99. Lotz JC, Colliou OK, Chin JR, Duncan NA, Liedenberg E. Compression-induced degeneration of the intervertebral disc: an in vivo mouse model and finite-element study. *Spine* 1998;23:2493-506.
100. Iatridis JC, Mente PL, Stokes IA, Aronsson DD, Alini M. Compression-induced changes in intervertebral disc properties in a rat tail model. *Spine* 1999;24:996-1002.
101. Kroeber M, Unglaub F, Guehring T, et al. Effects of controlled dynamic disc distraction on degenerated intervertebral discs: an in vivo study on the rabbit lumbar spine model. *Spine* 2005;30:181-7.
102. Hutton WC, Ganey TM, Elmer WA, et al. Does long-term compressive loading on the intervertebral disc cause degeneration? *Spine* 2000;25:2993-3004.
103. Hutton WC, Toribatake Y, Elmer WA, et al. The effect of compressive force applied to the intervertebral disc in vivo: a study of proteoglycans and collagen. *Spine* 1998;23:2524-37.
104. Handa T, Ishihara H, Ohshima H, et al. Effects of hydrostatic pressure on matrix synthesis and matrix metalloproteinase production in the human lumbar intervertebral disc. *Spine* 1997;22:1085-91.
105. Ishihara H, Urban JP. Effects of low oxygen concentration and metabolic inhibitors on proteoglycan and protein synthesis rates in the intervertebral disc. *J Orthop Res* 1999;17:829-35.
106. Ohshima H, Urban JPG. Effect of lactate concentrations and pH on matrix synthesis rates in the intervertebral disc. *Spine* 1992;17:1079-82.
107. Horner HA, Urban JP. Effect of nutrient supply on the viability of cells from the nucleus pulposus of the intervertebral disc. *Spine* 2001;26:2543-9.
108. Akmal M, Kesani A, Anand B, et al. Effect of nicotine on spinal disc cells: a cellular mechanism for disc degeneration. *Spine* 2004;29:568-75.
109. Uei H, Matsuzaki H, Oda H, et al. Gene expression changes in an early stage of intervertebral disc degeneration induced by passive cigarette smoking. *Spine* 2006;31:510-14.
110. Aufdermaur M, Fehr K, Lesker P, Silberberg R. Quantitative histochemical changes in intervertebral discs in diabetes. *Exp Cell Biol* 1980;48:89-94.
111. Robinson D, Mirovsky Y, Halperin N, Evron Z, Nevo Z. Changes in proteoglycans of intervertebral disc in diabetic patients: a possible cause of increased back pain. *Spine* 1998;23:849-56.
112. Stirling A, Worthington T, Rafiq M, Lambert PA, Elliott TS. Association between sciatica and Propionibacterium acnes. *The Lancet* 2001;357:2024-5.
113. Fritzell P, Bergström T, Welinder-Olsson C, et al. Detection of bacterial DNA in painful degenerated spinal discs in patients without signs of clinical infection. *Eur Spine J* 2004;13:702-6.
114. Sugimori K, Kawaguchi Y, Morita M, Kitajima I, Kimura T. High-sensitivity analysis of serum C-reactive protein in young patients with lumbar disc herniation. *J Bone Joint Surg [Br]* 2003;85-B:1151-4.
115. Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 1999;340:448-54.
116. Weinstein J. Mechanisms of spinal pain: the dorsal root ganglion and its role as a mediator of low-back pain. *Spine* 1986;11:999-1001.
117. Pedrini-Mille A, Weinstein JN, Found EM, Chung CB, Goel VK. Stimulation of dorsal root ganglia and degradation of rabbit annulus fibrosus. *Spine* 1990;15:1252-6.
118. Bogduk N, Twomey LT. *Clinical anatomy of the lumbar spine*. Second ed. Melbourne: Churchill Livingstone, 1991.
119. Gertzbein SD, Tile M, Gross A, Falk R. Autoimmunity in degenerative disc disease of the lumbar spine. *Orthop Clin North Am* 1975;6:67-73.
120. Takada T, Nishida K, Doita M, Kurosaka M. Fas ligand exists on intervertebral disc cells: a potential molecular mechanism for immune privilege of the disc. *Spine* 2002;27:1526-30.
121. Wang J, Tang T, Yang H, et al. The expression of Fas ligand on normal and stabbed-disc cells in a rabbit model of intervertebral disc degeneration: a possible pathogenesis. *J Neurosurg Spine* 2007;6:425-30.
122. Aihara T, Takahashi K, Ogasawara A, et al. Intervertebral disc degeneration associated with lumbosacral transitional vertebrae: a clinical and anatomical study. *J Bone Joint Surg [Br]* 2005;87-B:887-91.
123. Lee CK. Accelerated degeneration of the segment adjacent to a lumbar fusion. *Spine* 1988;13:375-7.
124. Phillips FM, Reuben J, Wetzell FT. Intervertebral disc degeneration adjacent to a lumbar fusion. *J Bone Joint Surg [Br]* 2002;84-B:289-94.
125. Panjabi MM, Andersson GB, Jorneus L, Hult E, Mattsson L. In vivo measurements of spinal column vibrations. *J Bone Joint Surg [Am]* 1986;68-A:695-702.
126. Wilder DG, Pope MH. Epidemiological and aetiological aspects of low back pain in vibration environments: an update. *Clin Biomech (Bristol, Avon)* 1996;11:61-73.
127. Wilder DG, Woodworth BB, Frymoyer JW, Pope MH. Vibration and the human spine. *Spine* 1982;7:243-54.
128. Pope MH, Kaigle AM, Magnusson M, Broman H, Hansson T. Intervertebral motion during vibration. *Proc Inst Mech Eng [H]* 1991;205:39-44.
129. Pope MH, Hansson TH. Vibration of the spine and low back pain. *Clin Orthop* 1992;279:49-59.
130. Bongers PM, Hulshof CT, Dijkstra L, et al. Back pain and exposure to whole body vibration in helicopter pilots. *Ergonomics* 1990;33:1007-26.

131. Frymoyer JW, Pope MH, Costanza MC, et al. Epidemiologic studies of low-back pain. *Spine* 1980;5:419-23.
132. Kelsey JL. An epidemiological study of acute herniated lumbar intervertebral discs. *Rheumatol Rehabil* 1975;14:144-59.
133. Kelsey JL. An epidemiological study of the relationship between occupations and acute herniated lumbar intervertebral discs. *Int J Epidemiol* 1975;4:197-205.
134. Bovenzi M, Zadini A. Self-reported low back symptoms in urban bus drivers exposed to whole-body vibration. *Spine* 1992;17:1048-59.
135. Kelsey JL, Hardy RJ. Driving of motor vehicles as a risk factor for acute herniated lumbar intervertebral disc. *Am J Epidemiol* 1975;102:63-73.
136. Heliövaara M. Occupation and risk of herniated lumbar intervertebral disk and sciatica leading to hospitalization. *J Chron Dis* 1987;40:259-64.
137. Jensen MV, Tüchsen F, Ørsted E. Prolapsed cervical intervertebral disc in male professional drivers in Denmark, 1981 - 90: a longitudinal study of hospitalizations. *Spine* 1996;21:2352-55.
138. Battisti MC, Videman T, Gibbons LE, et al. Occupational driving and lumbar disc degeneration: a case-control study. *Lancet* 2002;360:1369-74.
139. Kumar A, Varghese M, Mohan D, et al. Effect of whole-body vibration on the low back: a study of tractor-driving farmers in north India. *Spine* 1999;24:2506-15.
140. Videman T, Simonen R, Usenius J, Osterman K, Battisti M. The long-term effects of rally driving on spinal pathology. *Clin Biomech (Bristol, Avon)* 2000;15:83-6.
141. Drerup B, Granitzka M, Assheuer J, Zerlett G. Assessment of disc injury in subjects exposed to long-term whole-body vibration. *Eur Spine J* 1999;8:458-67.
142. Krismer M, Haid C, Rabi W. The contribution of annulus fibers to torque resistance. *Spine* 1996;21:2551-7.
143. Bogduk N, Twomey LT. *Clinical anatomy of the lumbar spine and sacrum*. Melbourne, Australia: Churchill Livingstone, 1987.
144. Farfan H. *Mechanical disorders of the low back*. Philadelphia: Lea & Febiger, 1973.
145. Liu YK, Goel VK, Dejong A, et al. Torsional fatigue of the lumbar intervertebral joints. *Spine* 1985;10:894-900.
146. Adams MA, Hutton WC. The mechanical function of the lumbar apophyseal joints. *Spine* 1983;8:327-30.
147. Farfan HF, Cossette JW, Robertson GH, Wells RV, Kraus H. The effects of torsion on the lumbar intervertebral joints: the role of torsion in the production of disc degeneration. *J Bone Joint Surg [Am]* 1970;52-A:468-97.
148. Farfan H. *The sciatic syndrome*. Thorofare: Slack Inc, 1996:75-121.
149. Hadjipavlou AG, Simmons JW, Yang JP, et al. Torsional injury resulting in disc degeneration. I: an in vivo rabbit model. *J Spinal Disord* 1998;11:312-17.
150. Sullivan JD, Farfan HF, Kahn DS. Pathologic changes with intervertebral joint rotational instability in the rabbit. *Can J Surg* 1971;14:71-9.
151. Ranson CA, Kerslake RW, Burnett AF, Batt ME, Abdi S. Magnetic resonance imaging of the lumbar spine in asymptomatic professional fast bowlers in cricket. *J Bone Joint Surg [Br]* 2005;87-B:1111-16.
152. Kasra M, Goel V, Martin J, et al. Effect of dynamic hydrostatic pressure on rabbit intervertebral disc cells. *J Orthop Res* 2003;21:597-603.
153. Ishihara H, McNally DS, Urban JP, Hall AC. Effects of hydrostatic pressure on matrix synthesis in different regions of the intervertebral disk. *J Appl Physiol* 1996;80:839-46.
154. Neidlinger-Wilke C, Würtz K, Urban JP, et al. Regulation of gene expression in intervertebral disc cells by low and high hydrostatic pressure. *Eur Spine J* 2006;15(Suppl 3):372-8.
155. Liu GZ, Ishihara H, Osada R, Kimura T, Tsuji H. Nitric oxide mediates the change of proteoglycan synthesis in the human lumbar intervertebral disc in response to hydrostatic pressure. *Spine* 2001;26:134-41.
156. Perek O. Fracture of the vertebral end-plate in the lumbar spine: an experimental biochemical investigation. *Acta Orthop Scand Suppl* 1957;25:1-101.
157. Brinckmann P, Biggemann M, Hilweg D. Prediction of the compressive strength of human lumbar vertebrae. *Spine* 1989;14:606-10.
158. Adams MA, McNally DS, Wagstaff J, Goodship AE. Abnormal stress concentrations in lumbar intervertebral discs following damage to the vertebral bodies: a cause of disc failure? *Eur Spine* 1993;1:214-21.
159. Adams MA, Freeman BJ, Morrison HP, Nelson IW, Dolan P. Mechanical initiation of intervertebral disc degeneration. *Spine* 2000;25:1625-36.
160. Hansson TH, Keller TS, Spengler DM. Mechanical behaviour of the human lumbar spine. II: fatigue strength during dynamic compressive loading. *J Orthop Res* 1987;5:479-87.
161. Rajasekaran S, Babu JN, Arun R, et al. A study of diffusion in human lumbar discs: a serial magnetic resonance imaging study documenting the influence of the endplate on diffusion in normal and degenerate discs. *Spine* 2004;29:2654-67.
162. Cinotti G, Della Rocca C, Romeo S, et al. Degenerative changes of porcine intervertebral disc induced by vertebral endplate injuries. *Spine* 2005;30:174-80.
163. Vernon-Roberts B, Fazzalari NL, Manthey BA. Pathogenesis of tears of the annulus investigated by multiple-level transaxial analysis of the T12-L1 disc. *Spine* 1997;22:2641-6.
164. Hamanishi C, Kawabata T, Yosii T, Tanaka S. Schmorl's nodes on magnetic resonance imaging: their incidence and clinical relevance. *Spine* 1994;19:450-3.
165. Hilton RC, Ball J, Benn RT. Vertebral end-plate lesions (Schmorl's nodes) in the dorsolumbar spine. *Ann Rheum Dis* 1976;35:127-32.
166. Paajanen H, Alanen A, Erkkälä M, Salminen JJ, Kivioja K. Disc degeneration in Scheuermann disease. *Skeletal Radiol* 1989;18:523-6.
167. Banerian KG, Wang AM, Samberg LC, Kerr HH, Wesolowski DP. Association of vertebral end plate fracture with pediatric lumbar intervertebral disk herniation: value of CT and MR imaging. *Radiology* 1990;177:763-5.
168. Goel VK, Monroe BT, Gilbertson LG, Brinckmann P. Interlaminar shear stresses and laminae separation in a disc: finite element analysis of L3-L4 motion segment subjected to axial compressive loads. *Spine* 1995;20:689-98.
169. MacLean JJ, Lee CR, Alini M, Iatridis JC. Anabolic and catabolic mRNA levels of the intervertebral disc vary with the magnitude and frequency of in vivo dynamic compression. *J Orthop Res* 2004;22:1193-200.
170. Chen J, Yan W, Setton LA. Static compression induces zonal-specific changes in gene expression for extracellular matrix and cytoskeletal proteins in intervertebral disc cells in vitro. *Matrix Biol* 2004;22:573-83.
171. Fujita K, Nakagawa T, Hirabayashi K, Nagai Y. Neutral proteinases in human intervertebral disc: role in degeneration and probable origin. *Spine* 1993;18:1766-73.
172. Holm S, Kaigle-Holm A, Ekstrom L, Karladani A, Hansson T. Experimental disc degeneration due to endplate injury. *J Spinal Disord Tech* 2004;17:64-71.
173. Chan D, Song Y, Sham P, Cheung KM. Genetics of disc degeneration. *Eur Spine J* 2006;15(Suppl 3):317-25.
174. Bass EC, Duncan NA, Hariharan JS, et al. Frozen storage affects the compressive creep behavior of the porcine intervertebral disc. *Spine* 1997;22:2867-76.
175. Johnstone B, Urban JP, Roberts S, Menage J. The fluid content of the human intervertebral disc: comparison between fluid content and swelling pressure profiles of discs removed at surgery and those taken postmortem. *Spine* 1992;17:412-16.